

Estimates of genetic selection gain in irrigated F₃ rice families

Estimativa do ganho de seleção em famílias F₃ de arroz irrigado

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Abstract

Rice production estimate in Brazil is 10.7 million tons, of which approximately 80% comes from the flood irrigation system in the southern region. Rice breeding programs have an important role in the development of cultivars with high production potential and industrial quality. This study estimates variance components and genetic parameters in F₃ families, and predicts the genetic gain in rice grain quality for the F₄ generation. We used fifty-six (56) F₃ families obtained from the advance of generations of the hybrid BRSCIRAD 302. This hybrid was selected for its traits of agronomic and scientific interest, such as grain yield and industrial yield. The hybrid was developed by Embrapa in partnership with CIRAD (Center for International Cooperation in Agronomic Research for Development) for Rio Grande do Sul State through the crossing of CIRAD 464 (M) × SBT 106 (F). The quality of the selected families was analyzed for amylose content, white belly, and gelatinization temperature. The F₃ families showed low gelatinization temperature. Selection of BRSCIRAD 302 hybrid families with smaller white belly spots and higher amylose content may be more successful if carried out between F₃ families. Twenty-two (22) families showed amylose content higher than that of the highest parent (29.44%). The predicted genetic gain in amylose content was 4.85% for families derived from the hybrid BRSCIRAD 302.

Additional keywords: genetic variation; grain quality; *Oryza sativa* L.

Resumo

O Brasil tem uma estimativa de produção de arroz de 10,7 milhões de toneladas e aproximadamente 80% é oriundo do sistema irrigado por inundação da região Sul do Brasil. Os programas de melhoramento genético de arroz têm importante papel no desenvolvimento de cultivares com elevado potencial produtivo e qualidade industrial. O objetivo deste trabalho foi estimar em famílias F₃ os componentes de variância, parâmetros genéticos e prever o ganho genético de seleção para geração F₄ sobre a qualidade de grãos em arroz. Para o presente estudo foram utilizadas 56 famílias F₃ obtidas com o avanço de geração do híbrido BRSCIRAD 302. O híbrido BRSCIRAD 302 foi selecionado por apresentar caracteres de interesse agrônomo e científico, como rendimento de grãos e rendimento industrial. Desenvolvido pela Embrapa em parceria com o CIRAD (Centro de Cooperação Internacional em Pesquisa Agrônoma para o Desenvolvimento) para o Rio Grande do Sul através do cruzamento do CIRAD 464 (M) × SBT 106 (F). As famílias selecionadas foram submetidas a análises de qualidade: teor de amilose, centro branco e temperatura de gelatinização. As famílias F₃ apresentaram baixa temperatura de gelatinização. A seleção de famílias do híbrido BRSCIRAD 302 com menor centro branco e maior teor de amilose poderá ter maior êxito se realizada entre as famílias F₃. 22 famílias apresentam teor de amilose superior ao maior genitor (29,44%). O ganho genético estimado para o teor de amilose foi de 4,85% nas famílias derivadas do híbrido BRSCIRAD 302.

Palavras-chave adicionais: *Oryza sativa* L.; qualidade de grãos; variação genética.

Introduction

The rice crop has great social and economic importance and is considered a crop that can alleviate world hunger. It is the third most produced cereal in the world, being the main component of the basic food basket for more than half the population (Magalhães Júnior & Oliveira, 2008; Fonseca et al., 2018a; Streck et al., 2018a).

Rice is one of the most consumed foods in Brazil, being characterized within the Brazilian economy as a product of very inelastic demand, that is, its demand does not change substantially with the increase in price. Regarding rice cultivation in Brazil, especially in irrigated crops in Rio Grande do Sul State, the quality and quantity of the product are often impaired by abiotic stresses such as low temperatures (Fonseca et al., 2018b).

Quantity and quality are important aspects for all the agents involved in the production, commercialization, and consumption of the cereal. However, genetic breeders should focus on consumer demands, which are made possible by the producers and social agents linked to the processing facilities. The analysis of the genetic breeding program for irrigated rice at Embrapa Clima Temperado - Estação Terras Baixas showed significant annual genetic progress between 1972 and 2016 for the main attributes of grain quality. The program has provided cultivars that meet the national demand for grain quality in the various segments of the rice production chain (Streck et al., 2018a). In this sector, genetic breeding has successfully achieved desirable production traits and industrial quality (Streck et al., 2018b).

From a genetic point of view, the breeding and evolution of cultivated plants have many characteristics in common: they require genetic variation, a selection method, and a degree or form of isolation to preserve the traits that arise (Briggs & Knowles, 1999). Therefore, identification of variation sources, which is the basis of plant breeding (Streck et al., 2017), is of fundamental importance for progress in improving agronomic traits of interest.

The concept of quality is conceived and viewed differently depending on the purpose of consumption, the ethnic group involved, postharvest processing, among others (Xu et al., 2015). The demands of all segments of the chain must be considered in determining the quality of a cultivar. Thus, it is important to know the organoleptic and physicochemical characteristics of rice grain, its cooking and appearance properties, which define quality standards in the market.

Grain quality traits limit commercialization in consumer countries. Physical appearance, culinary and sensory properties, and nutritional value are fundamental in the adoption of new varieties. The classification of each grain quality parameter varies according to the culture and consumption habits (Magalhães Júnior et al., 2011). To meet market demand and requirements for quality standards, the main rice research institutions have been developing rice cultivars with high grain yield and high grain quality, a fact that can be seen in the genetic gain in grain quality in the study of Streck et al. (2018b).

Thus, this study estimates variance components and genetic parameters in F_3 families, and estimate the genetic selection gain in rice grain quality for the F_4 generation.

Material and methods

Plant Material

The present study used F_3 progenies from the advance of generations of the hybrid BRSCIRAD 302 (F_1). This hybrid comes from the crossing between CIRAD 464 (M) × SBT 106 (F_1), and was developed in a collaborative process between Embrapa (Temperate Climate) and CIRAD for the conditions of Rio Grande do

Sul State. The superiority of grain yield (10.2 t ha^{-1}), combined with wide adaptability and stability and high industrial performance, were fundamental traits for its selection in this study (Streck et al., 2018c).

Generation Advance

In the 2011/2012 crop year, 4,500 seeds of the rice hybrid (F_2) were sown in two seasons and two locations: an experimental field at Estação Terras Baixas, belonging to Embrapa Clima Temperado, in Capão do Leão city, Rio Grande do Sul State; and an experimental field of IRGA (Instituto Rio Grandense do Arroz), in Cachoeirinha city, Rio Grande do Sul State. One hundred (100) plants were selected from each season and location, totaling 400 plants (F_2) with F_3 seeds. More details are obtained in Fonseca et al. (2019).

Conduction of F_3

Seeds of 400 F_3 plants were sown in the 2012/2013 crop year in the experimental field of Embrapa Clima Temperado - Estação Terras Baixas. Sowing was performed on December 17 and seedlings germinated on December 28, 2012. The area was managed according to the technical recommendations for irrigated rice cultivation (Sosbai, 2014). One hundred and fifty-four (154) plants were selected from fifty-six (56) families. Plants were selected through visual observation, considering agronomic traits of interest such as: lodging resistance, plant architecture, plant height, panicle exertion, yield capacity, grain spot, grain size, spikelet sterility, tillering capacity, disease resistance.

Experimental Design

The experimental design used in the F_3 generation was that of families with interspersed parents. 56 families were used, where the parents were arranged in three interspersed replicates between the families. Six parent plants were evaluated within each replicate.

Evaluated Traits

The seeds of the selected plants were subjected to the following quality analyses: amylose content, white belly, and gelatinization temperature.

Amylose content analysis: amylose is the linear starch fraction found in nonglutinous cultivars. Amylose content was analyzed according to the methodology described by Ciat (Martinez, 1989), using 90 whole, polished, and healthy grains. The content is thus framed by the following criteria: sticky - 0 to 2% amylose; glutinous - 3 to 10% amylose; low - 11% to 22% amylose; intermediate - 23% to 27% amylose; and high - 28% to 32% amylose. Data were expressed in percentages (%).

Amylose content was measured using the iodine colorimetric method. After making a flour from the rice grains, 100 mg of sample was weighed and trans-

ferred to a volumetric flask. Then, 1 mL of 95% alcohol was added to the flask, with care to dilute the rice flour for avoiding lump formation. Subsequently, 9 mL of sodium was added to the solution, which was heated for 9 minutes in a water bath (100 °C) to gelatinize the starch.

After cooling for 30 minutes, the flask walls were washed with distilled water, making up the total volume, and the solution was stirred. From this solution, a 5 mL aliquot was placed in a 100 mL flask, to which 1 mL of 1N acetic acid and 2 mL of iodine solution were added. Then, the total volume was filled with distilled water, and the solution was stirred and left to rest for thirty minutes.

The iodine solution was prepared by dissolving 0.2 g of metal iodine and 2 g of potassium iodide in 100 mL of distilled water. The absorption of the solution was measured using a spectrophotometer. Amylose content was quantified from a standard curve generated by absorption values of three known rice cultivars: IRGA 417, Bluebelle, and Bico Torto, with high, intermediate, and low amylose content, respectively.

White belly rate was determined in samples of whole grains after breeding, with scores from 0 to 5, according to the methodology described by Ciat (Martinez, 1989). White belly rate data were transformed following: $Wb = \sqrt{x + 0.5}$, where Wb : white belly of the transformed data, and x : the value of the collected datum.

Gelatinization temperature: the gelatinization temperature of starch refers to the cooking temperature at which the water is absorbed and the starch granules increase irreversibly in size, with simultaneous loss of crystallinity. It is analyzed according to the methodology of Ciat (Martinez, 1989), using 10 whole and polished grains, whose temperature is classified as low - from 63 to 68 °C; intermediate - from 69 to 73 °C; and high - from 74 to 80 °C.

Statistical Analysis

Data were submitted to analysis of variance according to the experimental design. In this sequence, sums of squares (SS) and mean squares (MS) were obtained for each of the model components. Subsequently, variance components and genetic parameters were estimated for the variables amylose content and white belly. Finally, the selection differential (SD) and genetic gain (GG) were estimated with the inbreeding coefficient (F) set at 75%, since the grains harvested from F_3 plants were in the F_4 generation. The coefficient was obtained according to Falconer & Mackay (1996), considering the self-fertilization mating system.

$$F_t = \frac{1}{2} (1 + F_{t-1})$$

Wherein: F is the inbreeding coefficient and, t corresponds to the generation.

Since the families were evaluated in trials

without replicates, it was necessary to correct the data according to the environmental effects estimated from parental information, as indicated by Cruz et al. (2012).

$$V_{cx} = V_{0x} - \varepsilon_f + \left(\frac{p_f - p_x}{p_f - p_i} \right) (\varepsilon_f - \varepsilon_i)$$

Wherein:

V_{cx} = corrected value of the segregating line in row x ;
 V_{0x} = original value of the segregating line in row x ;
 ε_f = value of the environmental effect that occurs in parents who are in position f of the rows;
 p_i = initial extreme position occupied by the parents;
 p_f = final extreme position occupied by the parents;
 p_x = position occupied by the segregating row, whose value is being corrected;
 ε_i = value of the environmental effect that occurs in parents who are in position i of the rows.

Variance components were estimated according to the following equations:

Environmental variance between ($\hat{\sigma}_{E \text{ between}}^2$) and within ($\hat{\sigma}_{E \text{ within}}^2$) families:

$$\hat{\sigma}_{E \text{ between}}^2 = \frac{1}{2} \left(\frac{\hat{\sigma}_{E \text{ between G1}}^2}{n} + \frac{\hat{\sigma}_{E \text{ between G2}}^2}{n} \right)$$

$$\hat{\sigma}_{E \text{ within}}^2 = \frac{\hat{\sigma}_{E \text{ within G1}}^2 + \hat{\sigma}_{E \text{ within G2}}^2}{2}$$

Wherein: $\hat{\sigma}_{E \text{ within G1}}^2$ is the environmental variance of parent 1; $\hat{\sigma}_{E \text{ within G2}}^2$ is the environmental variance of parent 2; n is the number observations (plants) associated with each parent.

Phenotypic variance between progenies ($\hat{\sigma}_{P \text{ between}}^2$), within progenies ($\hat{\sigma}_{P \text{ within}}^2$), and total phenotypic variance ($\hat{\sigma}_{P \text{ total}}^2$):

$$\hat{\sigma}_{P \text{ between}}^2 = \frac{\hat{\sigma}_{P \text{ between progenies}}^2}{n}$$

$$\hat{\sigma}_{P \text{ within}}^2 = \hat{\sigma}_{\text{within progenies}}^2$$

$$\hat{\sigma}_{P \text{ total}}^2 = \hat{\sigma}_{P \text{ within progenies}}^2 + \hat{\sigma}_{P \text{ between progenies}}^2$$

Genetic variance between ($\hat{\sigma}_{G \text{ between}}^2$) and within ($\hat{\sigma}_{G \text{ within}}^2$) progenies:

$$\hat{\sigma}_{G \text{ between}}^2 = \hat{\sigma}_{P \text{ between}}^2 - \hat{\sigma}_{E \text{ between}}^2$$

$$\hat{\sigma}_{G \text{ within}}^2 = \hat{\sigma}_{P \text{ within}}^2 - \hat{\sigma}_{E \text{ within}}^2$$

Additive genetic variance ($\hat{\sigma}_A^2$):

$$\hat{\sigma}_A^2 = \left(\frac{\hat{\sigma}_{G \text{ between}}^2 - F \hat{\sigma}_{G \text{ within}}^2}{F(1 + F)} \right)$$

Wherein: F = inbreeding coefficient ($3/4$) = 0.75

Genetic dominance variance ($\hat{\sigma}_D^2$):

$$\hat{h}_{a\text{ between}}^2 = \left(\frac{\hat{\sigma}_G^2 \text{ between}}{\hat{\sigma}_P^2 \text{ between}} \right) \times 100$$

Broad-sense heritability between ($\hat{h}_{a\text{ between}}^2$) and within ($\hat{h}_{a\text{ within}}^2$) families:

$$\hat{h}_{a\text{ between}}^2 = \left(\frac{\hat{\sigma}_G^2 \text{ between}}{\hat{\sigma}_P^2 \text{ between}} \right) \times 100$$

$$\hat{h}_{a\text{ within}}^2 = \left(\frac{\hat{\sigma}_G^2 \text{ within}}{\hat{\sigma}_P^2 \text{ within}} \right) \times 100$$

Narrow-sense heritability between ($\hat{h}_{r\text{ between}}^2$) and within ($\hat{h}_{r\text{ within}}^2$) families:

$$\hat{h}_{r\text{ between}}^2 = \left(\frac{2F\hat{\sigma}_A^2}{\hat{\sigma}_P^2 \text{ between}} \right) \times 100$$

$$\hat{h}_{r\text{ within}}^2 = \left(\frac{(1-F)\hat{\sigma}_A^2}{\hat{\sigma}_P^2 \text{ within}} \right) \times 100$$

Total narrow-sense heritability ($\hat{h}_{r\text{ total}}^2$):

$$\hat{h}_{r\text{ total}}^2 = \left(\frac{(1+F)\hat{\sigma}_A^2}{\hat{\sigma}_P^2 \text{ total}} \right) \times 100$$

Selection between and within families followed the methodology of Cruz et al. (2012), and the expressions used were:

Selection between families:

$$GG_{\text{between}} = \hat{h}_{r\text{ between}}^2 \times SD$$

GG_{between} = genetic gain between families;

$\hat{h}_{r\text{ between}}^2$ = narrow-sense heritability between families;

SD = selection differential

$$SD = \bar{x}_{fs} - \bar{x}_0$$

\bar{x}_{fs} = average of selected families;

\bar{x}_0 = original average of the selected population

Selection within families:

$$GG_{\text{within}} = \hat{h}_{r\text{ within}}^2 \times SD_m$$

GG_{within} = genetic gain within families;

$\hat{h}_{r\text{ within}}^2$ = narrow-sense heritability within families

$$SD_m = \frac{1}{s} \sum_{i=1}^s SD_i = \frac{1}{s} \sum_{i=1}^s (\bar{x}_{ind(i)} - \bar{x}_{fs(i)})$$

SD = selection differential;

S = number of selected families;

$\bar{x}_{ind(i)}$ = average of higher individuals within family i ;

$\bar{x}_{fs(i)}$ = average of the i -th selected family

Statistical analyses were performed using the GENES software (Cruz, 2013).

Results and discussions

The fifty-six (56) families showed similarly low gelatinization temperature, so this variable was discarded from the other analyses. In their study on genetic progress for quality traits, Streck et al. (2018a) state that most of the released cultivars have high amylose content and low gelatinization temperature, emphasizing that rice breeding programs have focused on grain quality, especially in the last decades.

Table 1 shows the result of the analysis of variance for white belly. There was no significance for parents (parent 1 and parent 2), and there was a significant effect between plots for F_3 . Knowledge of the distribution of variance between and within families is of fundamental importance since the success of a selection method will depend on the magnitude of the genetic variation available in the population/families (Ramalho et al., 2012).

Table 1 - Summary of the analysis of variance, sum of square and mean square of parent 1, parent 2 and generation F_3 for the variable white center.

Source of variation	Parent 1			Parent 2	
	FD	SS	MS	SS	MS
Between plots	2	0.001089	0.000544 ^{ns}	0.0000	0.00000 ^{ns}
Within plots	6	0.009900	0.001650	0.009900	0.00165
Total	8	0.010900	0.001363	0.009900	0.00124
Variance component between			0.000000		0.00000
Generation F_3					
Source of variation	FD	SS	MS		
Between plots	55	1.723087	0.031329**		
Within plots	98	0.726900	0.007417		
Total	153	2.449300	0.016008		
Variance component between			0.008699		

FD: freedom degrees, SS: sum of squares, MS: medium square, ^{ns}: no significant at 5% of error probability. * and **: significant at 5 and 1% of error probability by F test.

Table 2 - Summary of the analysis of variance, sum of squares and mean square of parent 1, parent 2 and generation F₃, for the variable amylose.

Source of variation	Parent 1			Parent 2	
	FD	SS	MS	SS	MS
Between plots	2	0.666667	0.333333 ^{ns}	0.222222	0.111111 ^{ns}
Within plots	6	3.333333	0.555556	2.000000	0.333333
Total	8	4.000000	0.500000	2.222222	0.277775
Variance component between			0.000000		0.000000
Generation F ₃					
Source of variation	FD	SS	MS		
Between plots	55	252.078993	4.583254**		
Within plots	98	138.336167	1.411594		
Total	153	390.414300	2.551727		
Variance component between			1.153851		

FD: freedom degrees, SS: sum of squares, MS: medium square, ^{ns}: no significant at 5% of error probability. * and **: significant at 5 and 1% of error probability by F test.

Analysis of variance for the variable amylose (Table 2) showed no significance between plots for both parents (1 and 2). On the other hand, the analysis between families showed a significant effect, which points to genetic variability between F₃ families.

Table 3 shows the estimates of the variance components and genetic parameters of the variables white belly and amylose content.

The phenotypic variance for the trait white belly is higher between families than within families. However, knowing the importance of phenotypic variance, the plant breeder is guided by the causes of genetic variations for selection purposes (Falconer & Mackay, 1996). According to Magalhães Júnior et al. (2011), recent changes in the classification of rice indicate that white-belly grains can be considered as plastered if grain opacity exceeds 50%, depreciating the product. Although white belly has several origins, such as early harvest and out-of-season heat waves, the genetic issue is present, and genotypes with a low incidence of white belly should be sought. Grain dimensions are also important, as the length/thickness ratio determines the rice class. The following classes are presented as a function of grain type: long-thin, long, medium, short, and mixed, among which the long-thin class stands out in the market.

Genetic variance between families for the traits white belly and amylose content was 0.011307 and 1.593316, respectively. Genetic variance within families was 0.005767 for white belly and 0.967149 for amylose content. As expected, for the two traits evaluated, estimates of genetic variance between families were higher than estimates within families. In addition to expressing genetic variation among individuals in a population, genotypic variance can be treated in the biometric context and be subdivided into causes of variation (additive, dominance, and epistatic). These are important components for the establishment of selection strategies in breeding programs.

Additive genetic variance was 0.005319 for white belly and 0.661298 for amylose (Table 3). This variance represents the inheritable fraction of the trait. Only the phenotypic value of an individual can be measured; however, it is the genetic value that will influence the next generation. Therefore, it is important

to know how much of the phenotypic variation is attributed to the genotypic variation, and this is measured by heritability (Falconer & Mackay, 1996; Cruz, 2005).

Amylose is considered the main factor related to the sensory quality of rice. Grain amylose content can be classified as: reduced content, for grains with 10 to 20% amylose; medium content, from 20 to 25%; and high content, from 25 to 33%. Grains can also be grouped into waxy rice - with 0 to 2% amylose; Japanese rice (short grain) - 18 to 20%; Indian rice - 25 to 30%; three rice - 19 to 20%; basmati rice - 25 to 27%; and jasmine rice - 22 to 25% (Botelho et al., 2009).

Amylose content is related to the textural properties of rice, such as softness, cohesion, color, brightness, and expansion volume, providing information about the changes that occur during cooking. Rice with high amylose content usually presents dry and loose grains, which can harden after cooling. Low amylose cultivars have soft, watery, and sticky grains when cooking. Cultivars with intermediate content have dry, loose, and soft grains, even after cooling (Magalhães Júnior et al., 2011).

Dominance variation for the traits white belly and amylose was 0.017750 and 3.207298, respectively. The dominance variance in the traits revealed that the selection would be less effective in the initial generation, with high variance values. Dominance deviations are not likely to be maintained in advanced breeding generations.

Estimates of broad-sense heritability between families for the traits white belly and amylose were high: 0.992039 and 0.955575, respectively. For broad-sense heritability within families, these estimates were 0.777548 and 0.6885147, respectively. Estimates of broad-sense heritability between and within families were considerably high. However, these estimates may be inflated by dominance deviations, since all genetic variance (additive and dominance deviations) is considered. Heritability estimates express the confidence of the phenotypic value as a guide for the genotypic value, and are part of most of the expressions used in genetic breeding, mainly in the prediction of genetic gains from selection (Falconer & Mackay, 1996; Cruz, 2005).

Table 3 - Estimates of variance components and genetic parameters for white center characteristics and amylose content in F₃ families of irrigated rice.

Parameters	White center	Amylose content
Phenotypic variance between	0.011397	1.667390
Phenotypic variance within	0.007417	1.411594
Environmental variance between	0.000091	0.074074
Environmental variance within	0.001650	0.444444
Genetic variance between	0.011307	1.593316
Genetic variance within	0.005767	0.967149
Broad-sense heritability between	0.992039	0.955575
Broad-sense heritability within	0.777548	0.685147
Additive genetic variance	0.005319	0.661298
Genetic dominance variance	0.017750	3.207298
Narrow-sense heritability between	0.700026	0.594910
Narrow-sense heritability within	0.179276	0.117119
Total narrow-sense heritability	0.494731	0.375862

For broad-sense heritability, the estimates for the two variables studied were high, indicating that most of the variations obtained in these populations are of genetic origin. According to Stansfield (1974), heritability values greater than 0.5 are considered high, and values between 0.2 and 0.5 and less than 0.2 are considered medium and low, respectively.

Narrow-sense heritability is more useful because it quantifies the relative importance of the additive proportion of the genetic variance, which can be passed on to the next generation. The important thing in the evaluation of heritability is to know how much of the selection differential is expected to be realized in the next generation (Falconer & Mackay, 1996).

For total narrow-sense heritability, estimates were 0.494731 for white belly and 0.375862 for amylose. When narrow-sense heritability is high, selection in the early generations of self-fertilization is effective. On the other hand, with a low value, selection should only be practiced in the more advanced breeding generations, since the increase in homozygosity, a consequence of self-fertilization, increases narrow-sense heritability, especially within families (Robinson, 1963; Falconer & Mackay, 1996)

For narrow-sense heritability between families, estimates were 0.700026 and 0.594910 for white belly and amylose, respectively. For narrow-sense heritability within families, these estimates were 0.179276 and 0.117119, respectively. The estimates indicated that the selection between families would be more effective in the F₃ generation, showing higher values. For amylose, families with the highest averages would be selected, excluding the selection within families and considering only the selection between families. Selection of families for the trait white belly would be based on the lowest averages of the families.

The traits white belly and amylose content are important parameters that serve as guidelines for plant

breeding, as they are agronomically important, being linked to consumer acceptability both by the visual aspect of the polished grain (white belly rate) or by the cooking aspect (amylose content). For rice commercialization, it is necessary to consider specific requirements of the various segments involved in the production and commercial processes of this cereal. These processes highlight the physical quality of the grain, expressed by processing yield, grain yield, size, and appearance (Mingotte et al., 2012). Another important physical property related to the quality of rice grains is translucency (Edwards et al., 2017). In many grains, translucency is interrupted by opaque areas in the endosperm, called white belly spots, plastered grains, or plastered areas (Smiderle & Dias, 2008).

Genetic studies show that amylose content is controlled by a major gene, but it is also influenced by the effect of gene dosage and by minor modifiers (Kumar & Khush, 1987). Furthermore, the environment can influence amylose content, mainly due to the high temperature during grain ripening (Hakata et al., 2012).

For amylose content, progenies with averages higher than that of the highest parent (29.44%) will be selected. The selection pressure used between and within families to select families with an average higher than that of the highest parent was 40%.

From the prediction of genetic gains from selection for amylose content, 22 F₃ families were selected, whose averages were higher than that of the highest parent (29.44%). The average of these 22 selected families was 29.57%. The average of the 56 families evaluated was 28.33% (Table 4). Streck et al. (2018a) evaluated genetic progress from 1972 to 2016 for quality traits and observed significant genetic gains: 0.20% per year, for percentage of whole grains after breeding; -1.38% per year, for percentage of plastered grains and white belly; -0.77% per year, for total plastered area; -0.08% per year, for total whiteness; and -0.82% per year, for nonvitreous whiteness of grains.

Table 4 - Gain estimation by selection aiming to increase the amylose content in the F₄ population of irrigated rice.

	Genetic gain between
Selection differential	1.246
Narrow-sense heritability between	0.595
Selection gain	0.741
Selection gain (%)	2.618
Average of the individuals selected (GG)	29.57
Average of the highest parent	29.44
Average of the families	28.33

The analysis of the results for the prediction of selection gain (Table 4) shows positive gains in amylose content for the F₄ generation of irrigated rice. Based on the observed results of narrow-sense heritability, selection in F₃ should be carried out between families, where the estimated gains from selection for F₄ would be 0.741 or 2.618% for amylose. Selection gains are not high for the trait amylose; however, the average of the parents used is high, meeting the standards of the rice market. In this sense, the expected gains in families overcome the values of parents, and even of the highest parent, who already has high amylose content. Although the expected genetic gains from selection are not so high, there was genetic gain for breeding purposes in obtaining families with higher amylose content.

Conclusions

Gelatinization temperature is low (63 to 68 °C) for all evaluated families.

The F₃ families show genetic variation for the traits white belly and amylose, which allows genetic gains from selection via genetic breeding.

Selection for white belly and amylose can be prioritized among F₃ families. Twenty-two (22) families have a higher average amylose content than the highest parent.

The estimated genetic gain for amylose content was 2.618% in families derived from the hybrid BRSCIRAD 302.

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References

Briggs FN, Knowles PF (1999) Introduction to plant breeding. New York: Reinhold Publishing Corporation, 426p.

Botelho RBA, Araújo HMC, Zandonadi RP, Ramos KL. (2009) Transformação dos alimentos: cereais e leguminosas. In: Araújo WMC, Montebello NP, Botelho RBA, Borgo LA (Org.). Alquimia dos Alimentos. v. 2. Brasília: Senac, p. 321-388.

Cruz CD (2005). Princípios de genética quantitativa. Viçosa: UFV, 2005. 349 p.

Cruz CD, Regazzi AJ, Carneiro PCS (2012) Modelos biométricos aplicados ao melhoramento genético. 4. ed. Viçosa: Ed. UFV, 514 p.

Cruz CD (2013) GENES – a software package for analysis in experimental statistics and quantitative genetics. Acta Scientiarum Agronomy 35 (3):271-276.

Edwards JD, Jackson AK, McClung AM (2017) Genetic architecture of grain chalk in rice and interactions with a low phytic acid locus. Field Crops Research 205:116-123.

Falconer DS, Mackay TFC (1996) Introduction to Quantitative Genetics. 4th ed. New York: Longman, 464p.

Fonseca GDM, Luz VK, Nardino M, Oliveira VF, Fonseca MDM, Scalco CB, Bandeira JM, Castilhos RMV, Magalhães Júnior AM, da Maia LC, Costa de Oliveira A (2018^a) Response of rice genotypes subjected to salt stress. Communications in Plant Sciences 8:90-99.

Fonseca GDM, Luz VK, Nardino M, Oliveira VF, Bandeira JM, Magalhães Júnior AM, Maia L. C, Costa de Oliveira A (2018^b) Characterization of herbicide tolerant rice genotypes under hydroponic culture. Communications in Plant Sciences 8:100-111.

Fonseca GDM, Nardino M, da Fonseca MDM, da Luz VK, de Magalhães Júnior AM, de Oliveira AC, da Maia LC (2019). Caracterização de populações F₂ de arroz irrigado visando a seleção para tolerância a baixas temperaturas. Magistra, 30: 123-133.

Hakata M, Kuroda M, Miyashita T, Yamaguchi T, Kojima M, Sakakibara H, Mitsui T, Yamakawa H (2012) Suppression of α -amylase genes improves quality of rice grain ripened under high temperature. Plant Biotechnology Journal 10:1110-1117.

Kumar I, Khush GS (1987) Genetic analysis of different amylose levels in rice. Crop Science, Madison 27:1167-1172.

- Magalhães Júnior A, Fagundes PR, Franco DF, Nunes CD, Moraes OP, Neves PCF, Rangel PHN, Moura Neto FP, Severo ACM, Fonseca GM, Streck E, Schumacher FP (2011) BRS Pampa: Nova cultivar de arroz irrigado para o RS. In: Congresso Brasileiro de Arroz Irrigado, VII., Camburiú, Anais... Camburiú, 156-159.
- Magalhães Júnior AM, Oliveira AC Arroz. In: Origem e evolução de plantas cultivadas. Barbieri RL, Stumpf ERT (Org.). Brasília, DF: Embrapa Informação Tecnológica, 2008 p.187-218.
- Martinez CY, Cuevas F (1989) Evaluación de la calidad culinária y molinera del arroz. Guía del estudio. Cali: CIAT, 75p.
- Mingotte FLC, Hanashiro RK, Fornasieri Filho D (2012) Características físico-químicas do grão de cultivares de arroz em função da adubação nitrogenada. Semina: Ciências Agrárias 33:2605-2618. Supl. 1.
- Ramalho MAP, Abreu AFB, Santos JB, Nunes JAR (2012) Aplicações da Genética Quantitativa no Melhoramento de Plantas Autógamas. 1ed. Lavras: ed. UFLA 522p.
- Robinson P (1963) Heritability: a second look. In: Hanson, W.D.; Robinson, H.F. Statistical Genetics and Plant Breeding. Washington: NAS-NCR 609-614.
- Smiderle OJ, Dias CTS (2008) Época de colheita e qualidade fisiológica de sementes em arroz irrigado (*Oryza sativa* L. cv. BRS Roraima). Pesquisa Agropecuária Tropical 38:188-194.
- Stansfield WD (1974) Genética. São Paulo: McGraw-Hill do Brasil, 958 p.
- SOSBAI (2014) Sociedade Sul Brasileira de Arroz Irrigado. Arroz irrigado: Recomendações técnicas da pesquisa para o Sul do Brasil. In: XXX Reunião Técnica da Cultura do Arroz Irrigado. Bento Gonçalves/RS. Anais. Santa Maria, RS, SOSBAI, 192p.
- Streck EA, Aguiar GA, Magalhães Júnior AM, Facchinello PHK, Oliveira AC (2017) Variabilidade fenotípica de genótipos de arroz irrigado via análise multivariada. Revista Ciência Agronômica 48:101-109.
- Streck EA, Magalhães Júnior AM, Aguiar GA, Facchinello PHK, Perin L, Fagundes PRR, Oliveira AC (2018a) Progresso genético da qualidade dos grãos de cultivares de arroz irrigado por inundação no Rio Grande do Sul. Pesquisa Agropecuária Brasileira 53(4):453-463.
- Streck EA, Magalhães AM, Aguiar GA, Facchinello PHK, Reis Fagundes PR, Franco DF, Nardino M, de Oliveira AC (2018b) Genetic Progress in 45 Years of Irrigated Rice Breeding in Southern Brazil. Crop Science 58: 1094-1105.
- Streck EA, Magalhães Júnior AM, Fagundes PRR, Aguiar GA, Facchinello PHK, Oliveira AC (2018c) Adaptability and stability of flood-irrigated rice cultivars released to the subtropical region of Brazil. Pesquisa Agropecuária Brasileira 53(4):1140-1149.
- Xu Q, Chen W, Xu Z (2015) Relationship between grain yield and quality in rice germplasms grown across different growing areas. Breeding Science 65: 226-232.